

## High-sensitivity C-reactive protein, Malondialdehyde and their association with Glycated hemoglobin (HbA1c) in type 2 diabetes patients

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### Abstract

**Background:** Evaluation of High-sensitivity C-reactive protein, malondialdehyde(MDA) levels in type 2 diabetic patients compare with healthy controls and correlate these levels with glycated hemoglobin (HbA1C) and insulin resistance. **Materials and Methods:** A prospective evaluation study was carried out in the Department of Biochemistry, Netaji Subhas Medical College and Hospital, Patna, Bihar India. The study population consisted of 200 subjects divided in to two groups viz., diabetic patients (type 2 diabetic subjects; n=100) and non- diabetic participants (n=100). 100 male and 100 female were include in this study. The age of the patients of both sex were 30-50 years. Serum hs- CRP and insulin was assessed by ELISA, malondialdehyde (MDA) was assessed by Thiobarbituric Acid Reactive Substances (TBARS) method and other routine investigations were carried out by standardized protocols with vitros 350 fully automated analyzer. **Results:** The mean serum hs-CRP and MDA levels were significantly high in type 2 diabetic patients compared with healthy patients. Hs-CRP and MDA levels we are shown significant positive correlation with glycosylated hemoglobin (HbA1C), insulin resistance, triglycerides and negative correlation with HDL cholesterol. **Conclusion:** Elevated hs - CRP, MDA levels are potentially important diagnostic markers for the assessment of endothelial dysfunction in type 2 diabetic patients. Tight blood glucose control, regular monitoring of hs-CRP, MDA levels within normal range might be useful for reduction of vascular complications in type 2 diabetic patients

**Keywords:** Abdominal malignancies, Incidence, Intestinal obstruction, Perforation.

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### Introduction

Diabetes mellitus (DM) considered as a widespread global disease. Conferring to recent reports, about 171 million persons in the world with DM in the year 2000 and this number expected to increase to 366 million through 2030. This disease is correlated with reducing life expectancy and significant other illnesses due to its relationship with microvascular complications (ischaemic heart disease, stroke and peripheral vascular disease), as a result led to lessen life quality[1]. Glycated hemoglobin (HbA1c) represents the blood glucose average level within the past 3 months.

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Therefore, HbA1c is a very important biochemical parameter that provide long term status of blood glucose levels and monitoring tool for measuring glycemic control in Type – 2 diabetic patients[2]. HbA1c in general, developed when the hemoglobin joined with glucose in the blood and become glycated[3]. According to many studies, HbA1c levels could be used as an independent risk factor for stroke and Cardiovascular disease (CVD) in both healthy and diabetics persons. It has been found that a (0.2%) decrease of HbA1c level can lower the risk of CVD development by 10%[4]. Furthermore, many studies have revealed, newborns moms with high HbA1c levels are more likely suffering from development of CVD in the future[5]. Chronic hyperglycemia and oxidative stress increases the pro-inflammatory proteins with infiltrated macrophages secreting inflammatory cytokines which leads to systemic

inflammation [6]. HsC reactive protein is an acute phase reactant protein produced by liver response to several cytokines and sensitive marker of low grade systemic inflammation[7,8]. Studies reported that hs -CRP directly binds to oxidized low-density lipoprotein cholesterol(LDL- C), induces plasminogen activator inhibitor-1 expression endothelial dysfunction by which leads to cardiovascular disease (CVD)[9-11]. Hyperglycemia induced oxidative stress induces pro inflammatory reactants with infiltrated macrophages secreting inflammatory cytokines which leads to local and systemic inflammation [12]. It has been recognized high levels of free radicals or reactive oxygen species (ROS), reactive nitrogen species (RNS) directly damage to the lipids which leads to formation of aldehydes such as malondialdehyde (MDA), propanal, hexanal, and 4-hydroxynonenal (4- HNE)[13]. So, in this view the objective of present study was to evaluate hs -CRP, MDA levels in type 2 diabetic patients and also to explore their association with HbA1c and insulin resistance.

#### Materials and methods

A prospective evaluation study was carried out in the Department of Biochemistry, Netaji Subhas Medical College and Hospital ,Patna, Bihar India from November 2019 to April 2020, after taking the approval of the protocol review committee and institutional ethics committee.

#### Study sample and inclusion/exclusion criteria

The study population consisted of 200 subjects divided in to two groups viz., diabetic patients (type 2 diabetic subjects; n=100) and non- diabetic participants (n=100). 100 male and 100 female were include in this

study. The age of the patients of both sex were 30-50 years. We excluded the patients on insulin, smokers, alcoholics, tobacco chewers, renal disease, inflammatory disorders, neoplastic disorders, thyroid disorders, liver dysfunction, and history of acute myocardial infarction, stroke and occlusive peripheral vascular disease.

#### Biochemical analysis

Fasting venous blood samples were collected from the study subjects and centrifuged at 3000 rpm for 15 min. Routine laboratory investigations were carried out by standardized protocols with vitros 350 fully automated analyzer. Serum insulin estimated by Enzyme Linked Immuno Sorbent Assay (ELISA), HbA1c estimated by (Ion Exchange Resin method) hs- CRP was assessed by (latex turbidimetric immunoassay), malondialdehyde (MDA) estimated by Thiobarbituric Acid Reactive Substances (TBARS) method[14]. Post prandial venous blood samples collected for plasma glucose (PPG) analysis. Homeostasis model assessment for Insulin Resistance (HOMA-IR) HOMA- IR calculated by using fasting glucose and insulin values:  $HOMA - IR = \text{fasting insulin} \times \text{fasting glucose} \text{ (mM/L)} / 22.51$ [15].

#### Statistical analysis

The recorded data was compiled and entered in a spreadsheet computer program (Microsoft Excel 2010) and then exported to data editor page of SPSS version 19 (SPSS Inc., Chicago, Illinois, USA). Descriptive statistics included computation of percentages and means. Test applied for the analysis was student t-test. The confidence interval and level of significance were set at 95% and 5%.

## Results

**Table 1: Gender and age distribution of patients**

Variables	N=200	Percentage %
Gender		
Male	100	50
Female	100	50
Age		
30-40 years	80	40
40-50 years	120	60

**Table 2: Comparison of baseline parameters in controls, type 2 diabetic patients**

Parameters	Controls(N=100)	T2DM(N=100)	p-value
Age	38.3±3.6	39.6±6.9	0.44 (NS)
Body mass index (BMI) kg/m <sup>2</sup>	25.1±1.3	28.1±2.6	0.012*
Waist/Hip ratio	0.92±0.02	0.94±0.11	0.024*
Systolic BP(mmHg)	116.3±5.7	119.2±9.4	0.08 (NS)
Diastolic BP (mm Hg)	77.1±5.8	80±7.8	0.13 (NS)

\*indicates statistical significance ( $\leq 0.05$ )

**Table 3: Comparison of FPG, PPG, HbA1C, HOMA-IR, Lipid profile, Liver profile, Renal profile hs-CRP and MDA levels in control and type 2 diabetic subjects**

Parameters	Controls (N=50)	T2DM(N=50)	p-value
FPG(mg/dl)	80.9± 8.7	134.0±13.5	0.021*
PPG(mg/dl)	106 .2±8.8	189±23 .6	0.032*
HbA1C	5.1±0.6	8.7±0.9	0.026*
Serum Triglycerides (mg/dl)	97.7±11.1	135.1±14.4	0.031*
Serumcholesterol (mg/dl)	179.8±9.8	207.2±22.8	0.042*
HOMA-IR	1.3±0.2	3.9±0.9	0.039*
HDLcholesterol (mg/dl)	44.0±1.9	40.1±3.1	0.027*
LDLcholesterol (mg/dl)	109±11.1	135.0±14.2	0.014*
Total Bilirubin(mg/dl)	0.76±0.06	0.78±0.07	0.72 (NS)
Direct Bilirubin(mg/dl)	0.2±0.07	0.19±0.08	0.41 (NS)
Serumurea(mg/dl)	22.9±4.6	27.2±6.9	0.24 (NS)
Serum creatinine(mg/dl)	0.67±0.3	0.78±0.7	0.311 (NS)
Hs-CRP(mg/L)	1.8±0.3	4.1±1.7	0.018*
MDA( $\mu$ mol/L)	1.8±0.6	5.9±1.6	0.029*

**Table 4: Correlation between hs-CRP & measured parameters in type 2 diabetic patients**

Parameters	Correlation Coefficient(r)
BMI	0.615**
W/H ratio	0.209
FBS	0.315*
PPBS	0.199
HbA1C	0.511**
HOMA-IR	0.482**
Cholesterol	0.249
TGL	0.301*
HDL	-0.333*
LDL	0.162
MDA	0.638**

\*Correlation is significant at the 0.05 level (2-tailed).

\*\*Correlation is significant at the 0.01 level (2-tailed)

**Table 5: Correlation between MDA & measured parameters in type 2 diabetic patient**

Parameters	Correlation Coefficient(r)
BMI	0.389**
W/Hratio	0.288*
FBS	0.609**
PPBS	0.191
HbA1C	0.415**
HOMA-IR	0.543**
Cholesterol	0.218
TGL	0.310*
HDL	-0.286*
LDL	0.121

\*Correlation is significant at the 0.05 level (2-tailed).

\*\*Correlation is significant at the 0.01 level (2-tailed)

## Discussion

Oxidative stress stimulates the inflammatory mediators which in turn enhances the production of reactive oxygen species (ROS) and reactive nitrogen species (RNS). Oxidative stress induces tumour necrosis factor alpha (TNF-  $\alpha$ ) secretion, it is linked to obesity related insulin resistance and vascular complications in type 2 diabetes mellitus[16,17]. Several studies explored that oxidative stress is not only due to free radical generation and also due to nonenzymatic protein glycosylation, auto-oxidation of glucose, impaired glutathione metabolism, decreased antioxidant capacity[18–20]. The present study has been shown significant increased hs-CRP and MDA levels in T2DM patients compared with healthy controls. Body mass index and (BMI) and Waist hip ratio were significantly increased in T2DM patients compared with healthy controls and also hs-CRP, MDA showed significant positive correlation with BMI. Obesity is considered as low-grade systemic inflammation, which results in metabolic derangements, insulin resistance and eventually precedes type 2 diabetes mellitus[21]. Obesity enhances sympathetic drive, increase vasomotor tone and hypertension; they proceed to metabolic abnormalities such as dyslipidemia, insulin resistance, inflammation, endothelial dysfunction and organ injury[22-24]. The present study also exhibits dyslipidemia in T2DM patients as reported earlier studies. High triglyceride levels and as well as decreased high-density lipoprotein (HDL) cholesterol, most likely underlying cause of increased free fatty acid flux, insulin resistance and vascular complications in type 2 diabetes mellitus[25,26]. We have observed significantly increased total cholesterol, triglycerides, LDL-C and decreased HDL-C in T2DM patients compared with healthy individuals and also hs-CRP, MDA levels were positively correlated triglycerides and negatively correlated with HDL cholesterol. In the present study we observed hs-CRP levels showed significant positive correlation with MDA, HbA1c and HOMA-IR. Chronic inflammation is potentially unifying mechanistic cause, accompanied by activation of major inflammatory pathways such as Jun-N-terminal kinases and the transcription factor NF-kappaB along with decreased HDL-cholesterol, with impairment in reverse cholesterol transport mechanism and parallel changes in apolipoproteins, enzymes, decreased anti-oxidant capacity[27-29]. Decreased HDL-Cholesterol and phospholipids could stimulate

accumulation of VLDL, which binds bacterial products and other toxic substances, resulting in hypertriglyceridemia.

Furthermore, it promotes lipid peroxidation by peroxynitrite formation by decreasing endogenous antioxidant defences and enhances the formation of atherosclerotic lesions[30]. ROS and RNS are collectively used to describe free radicals and other non-radical reactive derivatives known as oxidants. Biologically free radicals are highly unstable molecules which are products of normal cellular metabolism. Oxidative stress induced DNA damage markers such as 8-hydroxy-2'-deoxyguanosine (8-OHdG) and 8-oxo-7,8-dihydro-2'-deoxyguanosine; lipid-peroxidation products measured as thiobarbituric acid reactive substances (TBARS). In the present study we observed significantly increased MDA levels in T2DM patients compared to healthy controls and also positive correlation with HbA1c and HOMA-IR. HbA1c is widely used as mean glycemic index in diabetes and also useful measurement for the vascular complications. Oxidative stress plays a crucial role in pathogenesis of diabetic vascular complication[31]. Chronic hyperglycemia in diabetic patients can increase production of free radicals through Amadori rearrangement[32]. In general, the ROS and RNS are continuously generated in physiological conditions and are eliminated by several antioxidant enzymes.

Co-existence of inflammation, increased lipid peroxidation, dyslipidemia along with hyperglycemia conditions could pathologically increase the effect of oxidative stress[33]. However, the decreased efficiency of cellular antioxidant mechanisms with simultaneously enhanced lipid peroxidation along with increased insulin resistance and HbA1c may contribute factors of provoking inflammatory pathways and vascular complications in type 2 diabetes mellitus.

## Conclusion

Elevated hs-CRP, MDA levels are potentially important diagnostic markers for the assessment of endothelial dysfunction in type 2 diabetic patients. Tight blood glucose control, regular monitoring of hs-CRP, MDA levels within normal range might be useful for reduction of vascular complications in type 2 diabetic patients

## References

1. Ahmad, M., Ijaz, I., Rasheed, N., Saeed, M.,

- Ghaznavi, S., Mahmood, M., & Saleemi, A. Correlation between Glycated Hemoglobin and Dyslipidemia in Type-2 Diabetes Mellitus. *JIMDC*. 2016, 5(4):161-64.
2. Moinuddin K, Awanti SM. Evaluation of the relationship between glycemic parameters and serum uric acid level in type 2 diabetes mellitus patients, 2016;3:395-401.
  3. Jagtap MW, Rohankar PH, Kale SA. The Relation between serum uric acid & HbA1c in geriatric patients of Type 2 Diabetes in Amravati, Maharashtra, India. *Int. J. Bioassays*. 2016 ;5(06):4630-32.
  4. Arab, A. G., Zahedi, M., Nejad, V. K., Sanagoo, A., & Azimi, M. Correlation between Hemoglobin A1c and Serum Lipid Profile in Type 2 Diabetic Patients Referred to the Diabetes Clinic in Gorgan, Iran. *JCBR*, 2018;2(1):26-31.
  5. Gardiner HM, Pasquini L, Wolfenden J, Kulinskaya E, Li W, Henein M. Increased periconceptual maternal glycated haemoglobin in diabetic mothers reduces fetal long axis cardiac function. *Heart*. 2006; 92(8):1125-30.
  6. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. *J Clin Investig*. 2005; 115:1111-20.
  7. Ford ES. The metabolic syndrome and C-reactive protein, fibrinogen, and leukocyte count: findings from the Third National Health and Nutrition Examination Survey. *Atheroscler*. 2003; 168(2) :351-59.
  8. Pradhan AD, Manson JE, Rifai N, Buring JE, Ridker PM. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA*. 2001; 286(3):327-34.
  9. Devaraj S, Xu DY, Jialal I. C-Reactive Protein Increases Plasminogen Activator Inhibitor-1 Expression and Activity in Human Aortic Endothelial Cells. *Circulation*. 2003;107(3):398-404.
  10. Fichtlscherer S, Rosenberger G, Walter DH, Breuer S, Dimmeler S, et al. Elevated C-Reactive Protein Levels and Impaired Endothelial Vasoreactivity in Patients With Coronary Artery Disease. *Circ*. 2000;102(9):1000-06.
  11. Ridker PM, Buring JE, Cook NR, Rifai N. C-reactive protein, the metabolic syndrome, and risk of incident cardiovascular events. An 8-year follow-up of 14,719 initially healthy American women. *ACC Curr J Rev*. 2003;12(3):33-34.
  12. Wellen KE, Hotamisligil GS. Inflammation, stress, and diabetes. *J Clin Investig*. 2005; 115:1111-19.
  13. Moldovan L, Moldovan NI. Oxygen free radicals and redox biology of organelles. *Histochem Cell Biol*. 2004;122(4):395-412.
  14. Mahfouz MO, Hariprasad CH, Shaffie IA, Sadasivudu B. Serum malondialdehyde levels in myocardial infarction and chronic renal failure. *IRCS Med Sci*. 1986;14:1110-11.
  15. Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC. Homeostasis model assessment: insulin resistance and  $\beta$ -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia*. 1985; 28(7):412-19.
  16. Zhang P, Zhang X, Brown J, Vistisen D, Sicree R, et al. Global healthcare expenditure on diabetes for 2010 and 2030. *Diabetes Res Clin Pract*. 2010;87(3):293-301.
  17. Derosa G, D'Angelo A, Bonaventura A, Bianchi L, Romano D, et al. Effects of berberine on lipid profile in subjects with low cardiovascular risk. *Expert Opin Biol Ther*. 2013;13(4):475-82.
  18. Rosen P, Nawroth PP, King G, Mller W, Tritschler HJ, et al. The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes/Metab Res Rev*. 2001; 17(3): 189-212.
  19. Manna P, Jain SK. Obesity, Oxidative Stress, Adipose Tissue Dysfunction, and the Associated Health Risks: Causes and Therapeutic Strategies. *Metabolic Syndrome and Related Disorders*. 2015;13(10):423-44.
  20. Lee DH, Ha MH, Kim JH. Gamma-glutamyltransferase and diabetes: a 4 year follow-up study. *Diabetologia*. 2003;12:359-64.
  21. Gregor MF, Hotamisligil GS. Inflammatory Mechanisms in Obesity. *Ann Rev Immunol*. 2011;29(1):415-45.
  22. Visscher TL, Seidell JC. The Public Health Impact of Obesity. *Ann Rev Public Health*. 2001;22(1):355-75.
  23. Ouwens DM, Sell H, Greulich S, Eckel J. The role of epicardial and perivascular adipose tissue in the pathophysiology of cardiovascular disease. *J Cell Mol Med*. 2010;14(9):2223-34.
  24. Zhang H, Cui J, Zhang C. Emerging role of adipokines as mediators in atherosclerosis. *World J Cardiol*. 2010;2(11):370-76.

25. Jisieike-Onuigbo NN, Kalu OA, Onuigbo PC, Unuigbo EI, Oguejiofor CO. Prevalence of dyslipidemia among adult diabetic patients with overt diabetic nephropathy in Anambra state South-East Nigeria. *Nigerian J Clin Pract.* 2011;14(2):171-75.
26. Mooradian AD. Dyslipidemia in type 2 diabetes mellitus. *Nature Reviews Endocrinology.* 2009; 5(3):150-59.
27. Esteve E, Ricart W, Ferná'ndez-Real JM. Dyslipidemia and inflammation: an evolutionary conserved mechanism. *Clin Nutr.* 2005;24(1):16-31.
28. Hirosumi J, Tuncman G, Chang L, Go'rgu'n CZ, Uysal KT, et al. A central role for JNK in obesity and insulin resistance. *Nature.* 2002; 420(6913):333-336.
29. Nishimura S, Manabe I, Nagasaki M. CD8+ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nat Med.* 2009;15:914-34.
30. Mehdipour M, Zenouz AT, Davoodi F, Gholizadeh N, Damghani H, et al. Evaluation of the Relationship between Serum Lipid Profile and Oral Lichen Planus. *J Dent Res, Dent Clin, Dent Prospects.* 2015;9(4):261-66.
31. Schafer FQ, Buettner GR. Redox environment of the cell as viewed through the redox state of the glutathione disulfide/glutathione couple. *Free Radical Biol Med.* 2001;30(11):1191-12.
32. Giugliano D, Ceriello A, Paolisso G. Oxidative Stress and Diabetic Vascular Complications. *Diabetes Care.* 1996;19(3):257-67.
33. Pendyala G, Thomas B, Joshi S. Evaluation of total antioxidant capacity of saliva in type 2 diabetic patients with and without periodontal disease: A case-control study. *North Am J Med Sci.* 2013;5(1):51.

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