

Analysis of Haematological Parameters and Peripheral Smear findings of COVID- 19 Patient's Blood Sample

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Abstract

Introduction: Lymphopenia is an important finding in COVID-19 is a systemic infection which can help to predict the prognosis. The ratio of neutrophils to lymphocytes ratio, lymphopenia and thrombocytopenia can help to determine the severity of an incident. Severe thrombocytopenia can lead to potentially fatal consequences in the form of DIC. **Objectives:** To know the haematological parameters and peripheral smear findings in blood samples of COVID -19 patients and their relation with severity of infection. **Materials and Methods:** This prospective study was conducted at Department of Pathology, Hassan Institute of Medical Sciences, Hassan, Karnataka. 12288 confirmed cases from Covid -19 infection from June 2020 to May 2021, were evaluated in detail for complete blood count analysis by Cell Dyn Ruby and peripheral smear study. Findings were noted and correlated with severity of infection. **Results:** In the present study from asymptomatic cases to critical cases, there was gradual increase in leucocytes count, along with neutrophilia, absolute neutrophils count, neutrophil to lymphocyte ratio and gradual decrease in lymphocytes, absolute lymphocyte count and platelets. In our study, peripheral smear findings showed toxic granules along with other reactive changes in neutrophils. Lymphopenia worsens as the disease progresses to a more advanced state. Individual proper management and the disease's outcome can be monitored effectively by noting above discussed haematological parameters in COVID-19 infection.

Keywords: COVID-19 infection, Lymphopenia, Neutrophils to lymphocytes ratio, Thrombocytopenia.

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Introduction

COVID-19 is a systemic infection that has a significant impact on the hematopoietic system and hemostasis. Lymphopenia is an important test finding that can help to predict the prognosis. The ratio of neutrophils to lymphocytes, as well as the peak platelet/lymphocyte ratio, can help to determine the severity of an incident. Longitudinal monitoring of lymphocyte count dynamics, inflammatory indices, and other biochemical markers over the course of the disease may help in the identification of patients with poor prognoses and prompt action to improve outcomes. Severe thrombocytopenia can lead to potentially fatal consequences. The disorder known as disseminated intravascular coagulation (DIC) needs ongoing monitoring and therapy.

Aims and objectives of the study

To know the haematological parameters and peripheral smear findings in blood samples of COVID -19 patients and to correlate findings with severity of infection

Materials

Source of data

Blood samples received from the confirmed cases of Covid-19 patients.

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Estimation of sample size

Covid blood samples received from June 2020 to May 2021

Inclusion criteria

Blood samples of Covid patients.

Exclusion criteria

Blood samples of Non Covid patients

Methodology

Above prospective study was conducted from June 2020 to May 2021 at Department of Pathology, Hassan Institute of Medical Sciences, Hassan, Karnataka. Blood samples were collected from confirmed cases of Covid-19 patients. Samples were subjected for complete blood count analysis by Cell Dyn Ruby five-part automated blood sample analyser and peripheral smear study. Findings were noted and correlated with clinical symptoms and signs.

Results

Our prospective study period was from June 01, 2020 to May 2021. The age group ranged from 01 year to 90 years. The total number of blood samples subjected for investigations for coagulation abnormalities in the Department of Pathology laboratory of Hassan Institute of Medical Sciences, Hassan were 12288. The maximum number of samples were seen in age group of 41 to 50 years (19.31%). For male it was in 41 to 50 years (20.38%) and for female it was in 31 to 40 years (19.51%). [Table 01]

Table 1: Showing distribution of COVID-19 blood sample with respect to age and sex.

Age (In Years)	Male - Number of COVID-19 blood sample	Female - Number of COVID-19 blood sample	Total
01-10	237	204	441
11-20	618	579	1197
21-30	1296	894	2190
31-40	1326	1008	2334
41-50	1452	921	2373
51-60	1311	825	2136
61-70	642	522	1164
71-80	222	204	426
81-90	18	9	27
Total Number	7122	5166	12288

On receiving fresh, adequate COVID-19 blood samples to the Department of Pathology laboratory, samples were classified as below based on patient history, and categorization by clinicians.

1. Asymptomatic cases (AS): Patients' blood samples without any symptoms and signs where they had history of contacts with primary COVID-19 cases or coming on their own.
2. Influenza like illness (ILI)- Mild cases: Patient with history of fever, running nose, sore throat, cough with or without sputum production and other mild symptoms related to corona illness
3. Severe acute respiratory illness (SARI)- Moderate cases: Patient with high grade fever, chills, dyspnea, severe cough with expectoration and other symptoms of corona which impaired the daily routine life activities.
4. Critical cases (CC)- Severe cases: Critical illness was defined throughout the study as a requirement for endotracheal intubation, mechanical ventilation and careful monitoring and supervision by attending doctors.

On detailed analysis, of 4096 blood samples [Table 02] the contribution of Asymptomatic cases (AS), Influenza like illness (ILI), Severe acute respiratory illness (SARI) and Critical cases (CC) were 3252 (26.46%), 5286 (43.01%), 2973 (24.19%) and 777 (6.32%) respectively. Maximum number of asymptomatic cases (22.69%) were seen in 31 to 40 years and no cases in 81 to 90 years of age group. ILI cases were more commonly seen in 31 to 40 years (19.46%) and very minimal case (03 case) in 81 to 90 years of age group. SARI cases were more common in 41 to 50 years (21.08%) and only six cases from 81 to 90 years of age group. Maximum number of CC (28.57%) were seen in 61 to 70 years and no cases in 01 to 20 years of age group. Most of the critical cases were having history of chronic lung disease, cardiovascular disease, immunocompromised status, diabetes mellitus, chronic kidney disease requiring dialysis, chronic liver disease, and residence in a long-term care facility.

Table 02: Classification and typing of blood samples with respect to age.

Age (In Years)	Asymptomatic	Influenza like illness	Severe acute respiratory illness	Critical cases	Total
01-10	159	219	63	00	441
11-20	366	687	144	00	1197
21-30	711	984	486	09	2190
31-40	738	1029	519	48	2334
41-50	678	891	627	177	2373
51-60	423	915	603	195	2136
61-70	171	444	327	222	1164
71-80	06	114	198	108	426
81-90	00	3	6	18	27
Total Number	3252	5286	2973	777	12288

For all the received fresh COVID-19 blood samples in Pathology laboratory the following tests were performed according to standard procedure for coagulation abnormality. For each tests, control samples results were also noted.

1. Total Leukocyte count (TLC)
2. Absolute Neutrophil count (ANC)
3. Absolute Lymphocyte count (ALC)
4. Neutrophil count to Lymphocyte count ratio (NLR)
5. Platelet count (PC)

After subjecting the blood samples of 12288 COVID-19 positive cases, results were correlated with the type of blood samples and correlated with severity of infection.

Table 03: Showing the average values of TLC, ANC, ALC, NLR & PC of 12288 blood samples.

Sl No	Type of sample	TLC (Cells/cumm)	ANC (Cells/cumm)	ALC (Cells/cumm)	NLR	PC (lakhs Cells/cumm)
1	Control	7.42	5.60	2.73	2.05	2.93
2	AS	7.02	5.18	2.21	2.34	1.85
3	ILI	10.01	7.13	1.76	4.05	1.13
4	SARI	13.69	8.26	1.05	7.86	0.74
5	CC	21.85	9.04	0.74	12.21	0.39

Discussions

Coronavirus illness 2019 (COVID-19) is a respiratory tract infection caused by SARS-CoV-2, a newly emerged coronavirus initially identified in Wuhan, China, in December 2019. The virus's genetic

sequence implies that SARS-CoV-2 is a beta-coronavirus closely related to SARS[1].

While the majority of COVID-19 patients have a mild or uncomplicated sickness, about 14% develop severe disease that need hospitalisation and oxygen assistance, and 5% require admission to an

intensive care unit. SARS-CoV-2 infection has been recorded in individuals who never develop symptoms (asymptomatic) and in patients who are not yet symptomatic in several studies (pre-symptomatic)[2-6].

Males were affected more frequently than females, according to the demographic parameters of our study. This is comparable to studies by Guan WJ et al[7], and Nanshan Chen et al[8], in which disease incidence was higher in men than in women. Immunological, cultural, or racial differences in the Indian population might explain the shift in gender patterns. More research is needed in this area, and a bigger cohort study may be recommended to prove the point[1].

Lippi G et al[9] conducted a systematic evaluation of the literature and found that the most important leucocytosis, neutrophilia, lymphopenia and other haematological abnormalities seen in COVID-19 patients may predict the progression to severe or critical forms of COVID-19[1]. In the present study there is gradual increase in leucocytes count from asymptomatic cases to critical cases (7.02 to 21.85 cells/ cumm). As per the IFCC (Information Guide on COVID-19)[1] guidelines, the CBC parameters in a COVID case show neutrophilia, leucocytosis lymphopenia, and thrombocytopenia. Chuan Qin et al[10], found that primary dysregulation of the immune response, particularly T lymphocytes, may play a significant role in the pathogenesis of COVID-19.

In the present study there is gradual increase in absolute neutrophil count (ANC) count from asymptomatic cases to critical cases 5.18 to 9.04 cells/ cumm. A series of COVID-19 reports revealed that ICU patients were more likely to develop neutrophilia, a disease-progression sign. Various studies[11-13] have shown the median absolute neutrophil count (ANC) in ICU cases was 10.6 (5.0–11.8) 10⁹/L, substantially higher than the 4.4 (2.0–6.1) 10⁹/L in non-ICU cases. According to a retrospective study of COVID-19 patients' initial laboratory indices, 34.5 percent of them had neutrophilia, and patients with ARDS had more neutrophils than those who did not have ARDS. Furthermore, patients with ARDS who died had greater neutrophil counts than those who survived[14]. In the present study there is gradual decrease in absolute lymphocyte count (ALC) count from asymptomatic cases to critical cases (5.18 to 9.04 cells/ cumm). According to study done in Zhongnan Hospital of Wuhan University, discovered that lymphocytes, the principal antiviral cells, were prone to decrease continuously and severely in ICU and dead patients[12]. Lymphopenia is very commonly seen in this viral infection. Lymphopenia was found in 83.2 percent of patients on admission in a multicentre trial involving 1,099 patients from 552 locations in China. Many other studies in China found lymphopenia rates ranging from 26 to 80 percent[7,15]. T lymphocytes and NK cells were found to be depleted in COVID-19 patients[16,17,19]. The low cycle threshold value in respiratory samples indicated that lymphopenia on presentation was associated with a high viral load[20]. Liu et al[21], looked into the relationship between dynamic variations in nasopharyngeal virus load and lymphocyte count. In our study there is significant rise in NLR ratio in critical care patients (12.21). The patient's response to inflammatory insult may be indicated by a high NLR, with neutrophils increasing in reaction to stress, which, when overwhelming, causes lymphocyte apoptosis[22-25]. The Absolute Neutrophil to Lymphocyte Ratio (NLR) is a biomarker used to determine the severity of bacterial infections as well as the prognosis of pneumonia and tumour patients. NLR could be used as a surrogate marker for detecting COVID 19 infection early[1]. Normal NLR values in an adult, non-geriatric population in excellent health were found to be between 0.78 and 3.53 in a research by Forget et al[26]. In contrast, Jingyuan Liu[27] determined that patients over or equal to age 50 years with NLR of more than 3.53 are at risk of serious disease and should be admitted to the intensive care unit as soon as possible if necessary. A substantial percentage of cases in our study had toxic granules with other reactive changes and reactive lymphocytes, which matched the findings of Bingwen Eugene Fan et al[16]. In the present study there is decrease platelet counts from asymptomatic cases to critical cases (1.85 to 0.39 Lakhs cells/ cumm). In comparison to lymphopenia, thrombocytopenia is less common in patients with

COVID-19[15]. Previous SARS-CoV-1 patients' experiences suggested that coronavirus could cause thrombocytopenia through direct viral infection of bone marrow haematopoietic stem cells via CD13 or CD66a, the formation of autoantibodies and immune complexes, disseminated intravascular coagulopathy (DIC), and platelet consumption in lung epithelium[28,29]. SARS patients had higher levels of soluble vascular cell adhesion molecule-1 (sVCAM-1), which increased vascular sequestration and caused thrombocytopenia[30]. Some illnesses, such as infectious mononucleosis, Bordetella pertussis, and hantavirus, are characterised by atypical/reactive cells in the peripheral smear[24-26]. Infectious mononucleosis is characterised by a wide range of reactive/pleomorphic lymphocytes, whereas hantavirus is characterised by Downey type III cells or immunoblasts. These aren't specific results, and they could appear in different numbers in other infections, autoimmune disorders, and cancers[31-34].

Conclusions

In conclusion, the COVID-19 infection has caused significant haematological complications. In follow-up cases with persistent symptoms, a statistically significant increase in CBC parameters, NLR, was observed; however, a larger follow-up cohort is required to reach statistical significance.

Lymphopenia worsens as the disease progresses to a more advanced state. In patients with SARS-CoV-2 infection, increased neutrophil count and neutrophil-to-lymphocyte ratio, as well as lower haemoglobin concentration, have been identified as risk factors for severe sickness. Individual treatment decisions should be made to reduce the risk of infection while not risking the disease's outcome.

References

1. Neema Tiwari, Devajit Nath et al. The Neutrophil Lymphocyte Ratio (NLR), Platelet Lymphocyte Ratio (PLR) and routine hematological parameters of COVID-19 Patient: A perspective of the Indian scenario from a frontline pilot study of 32 COVID-19 cases in a Tertiary Care Institute of North India. MEDRXIV/2020/102913
2. Chan, J. F.-W., Yuan, S., Kok, K.-H., To, K. K.-W., Chu, H., Yang, J., Xing, F., Liu, J., Yip, C. C.-Y., and Poon, R. W.-S. (2020) A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: a study of a family cluster, *The Lancet* 395, 514-523.
3. Hoehl, S., Rabenau, H., Berger, A., Kortenbusch, M., Cinatl, J., Bojkova, D., Behrens, P., Böddinghaus, B., Götsch, U., and Naujoks, F. (2020) Evidence of SARS-CoV-2 infection in returning travelers from Wuhan, China, *New England Journal of Medicine* 382, 1278-1280.
4. Hu, Z., Song, C., Xu, C., Jin, G., Chen, Y., Xu, X., Ma, H., Chen, W., Lin, Y., and Zheng, Y. (2020) Clinical characteristics of 24 asymptomatic infections with COVID-19 screened among close contacts in Nanjing, China, *Science China Life Sciences*, 1-6.
5. Surveillances, V. The epidemiological characteristics of an outbreak of 2019 novel coronavirus diseases (COVID-19)—China, 2020, *China CDC Weekly*. 2020; 113-122.
6. Wang, Y., Liu, Y., Liu, L., Wang, X., Luo, N., and Ling, L. Clinical outcome of 55 asymptomatic cases at the time of hospital admission infected with SARS-Coronavirus-2 in Shenzhen, China, *The Journal of infectious diseases*. 2020:1
7. Guan, W.-j., Ni, Z.-y., Hu, Y., Liang, W.-h., Ou, C.-q., He, J.-x., Liu, L., Shan, H., Lei, C.-l., and Hui, D. S. (2020) Clinical characteristics of coronavirus disease 2019 in China, *New England Journal of Medicine*. 2020:9
8. Chen, N., Zhou, M., Dong, X., Qu, J., Gong, F., Han, Y., Qiu, Y., Wang, J., Liu, Y., and Wei, Y. Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study, *The Lancet* 2020;395, 507-513.

9. Lippi, G., and Plebani, M. (2020) Procalcitonin in patients with severe coronavirus disease 2019 (COVID-19): A meta-analysis, *Clinica Chimica Acta; International Journal of Clinical Chemistry*.
10. Qin, C., Zhou, L., Hu, Z., Zhang, S., Yang, S., Tao, Y., Xie, C., Ma, K., Shang, K., and Wang, W. (2020) Dysregulation of immune response in patients with COVID-19 in Wuhan, China, *Clinical Infectious Diseases*.
11. Huang CL, Wang YM, Li XW, Ren LL, Zhao JP, Hu Y, Zhang L, Fan G, Xu J, Gu X, Cheng Z, Yu T, Xia J, Wei Y, Wu W, Xie X, Yin W, Li H, Liu M, Xiao Y, Gao H, Guo L, Xie J, Wang G, Jiang R, Gao Z, Jin Q, Wang J, Cao B (2020) Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 395(10223):497–506
12. Wang DW, Hu B, Hu C, Zhu FF, Liu X, Zhang J, Wang B, Xiang H, Cheng Z, Xiong Y, Zhao Y, Li Y, Wang X, Peng Z (2020) Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus-infected pneumonia in Wuhan, China. *JAMA* 323(11):1061–1069
13. Liu J-Y, Liu Y, Xiang P, Pu L, Xiong H-F, Li C-S, et al. (2020) Neutrophil-to-lymphocyte ratio predicts severe illness patients with 2019 novel coronavirus in the early stage. medRxiv. <https://doi.org/10.1101/2020.02.10.20021584>
14. Xiaoqing Liu & Run Zhang & Guangsheng He; Hematological findings in coronavirus disease 2019: indications of progression of disease. *Annals of Hematology* (2020) 99:1421–1428
15. Carmen Ka Man Cheung, Man Fai Law, Grace Chung Yan Lui, Sunny Hei Wong, Raymond Siu Ming Wong; *Acta Haematol*. DOI: 10.1159/000510178
16. Fan BE, Chong VC, Chan SS, Lim GH, Lim KG, Tan GB, et al. Hematologic parameters in patients with COVID-19 infection. *Am J Hematol*. 2020 Jun; 95(6):E131–4.
17. Qin C, Zhou L, Hu Z, Zhang S, Yang S, Tao Y, et al. Dysregulation of immune response in patients with COVID-19 in Wuhan, China. *Clin Infect Dis*. 2020 Mar 12;ciaa248
18. Chen G, Wu D, Guo W, Cao Y, Huang D, Wang H, et al. Clinical and immunological features of severe and moderate coronavirus disease 2019. *J Clin Invest*. 2020 May; 130(5):2620–9.
19. Liu Z, Long W, Tu M, Chen S, Huang Y, Wang S, et al. Lymphocyte subset (CD4+, CD8+) counts reflect the severity of infection and predict the clinical outcomes in patients with COVID-19. *J Infect*. 2020 Apr; S0163-4453(20)30182-1.
20. Liu Y, Yang Y, Zhang C, Huang F, Wang F, Yuan J, et al. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Sci China Life Sci*. 2020 Mar; 63(3): 364–74.
21. Liu Y, Liao W, Wan L, Xiang T, Zhang W. Correlation between Relative Nasopharyngeal Virus RNA Load and Lymphocyte Count Disease Severity in Patients with COVID-19. *Viral Immunol*. 2020 Apr; vim.2020.0062.
22. Zahorec R. Ratio of neutrophil to lymphocyte counts—rapid and simple parameter of systemic inflammation and stress in critically ill. *Bratisl Lek Listy*. 2001;102(1):5–14.
23. O'Mahony JB, Palder SB, Wood JJ, McIrvine A, Rodrick ML, Demling RH, et al. Depression of cellular immunity after multiple trauma in the absence of sepsis. *J Trauma*. 1984 Oct;24(10):869–75
24. Dahn MS, Whitcomb MP, Lange MP, Jacobs LA. Altered T-lymphocyte subsets in severe sepsis. *Am Surg*. 1988 Jul;54(7):450–5.
25. Le Tulzo Y, Pangault C, Gacouin A, Guilloux V, Tribut O, Amiot L, et al. Early circulating lymphocyte apoptosis in human septic shock is associated with poor outcome. *Shock*. 2002 Dec;18(6):487–94.
26. Forget, P., Khalifa, C., Defour, J.-P., Latinne, D., Van Pel, M.-C., and De Kock, M. (2017) What is the normal value of the neutrophil-to-lymphocyte ratio?, *BMC research notes* 10, 12
27. Liu, J., Liu, Y., Xiang, P., Pu, L., Xiong, H., Li, C., Zhang, M., Tan, J., Xu, Y., and Song, R. (2020) Neutrophil-to-lymphocyte ratio predicts severe illness patients with 2019 novel coronavirus in the early stage, *MedRxiv*.
28. Yang M, Hon KL, Li K, Fok TF, Li CK. The effect of SARS coronavirus on blood system: its clinical findings and the pathophysiologic hypothesis. *Zhongguo Shi Yan Xue Ye Xue Za Zhi*. 2003; 11(3): 217–21.
29. Yang M, Ng MH, Li CK. Thrombocytopenia in patients with severe acute respiratory syndrome (review). *Hematology*. 2005 Apr; 10(2): 101–5.
30. Chen RF, Chang JC, Yeh WT, Lee CH, Liu JW, Eng HL, et al. Role of vascular cell adhesion molecules and leukocyte apoptosis in the lymphopenia and thrombocytopenia of patients with severe acute respiratory syndrome (SARS). *Microbes Infect*. 2006; 8(1): 122–7.
31. Alia Nazarullah, MD, Christine Liang, MD, Andrew Villarreal, MLS, Russell A Higgins, MD, Daniel D Mais, MD. *American Journal of Clinical Pathology*, aqa.2020;9
32. George TI. Malignant or benign leukocytosis. *Hematology Am Soc Hematol Educ Program*. 2012;2012:475–484
33. Klein E, Kis LL, Klein G. Epstein-Barr virus infection in humans: from harmless to life endangering virus-lymphocyte interactions. *Oncogene*. 2007;26(9):1297–05.
34. Koster F, Foucar K, Hjelle B, et al. Rapid presumptive diagnosis of hantavirus cardiopulmonary syndrome by peripheral blood smear review. *Am J Clin Pathol*. 2001;116:665–672

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