

A Study On Metallo-Beta Lactamase Producing Pseudomonas Aeruginosa Species In Clinical Isolates Of A Tertiary Care Hospital Of Bihar

Kumar Amaresh¹, Sagar Kumar^{2*}, Rakesh Kumar³

¹Assistant Professor, Department of Microbiology, JLNMCH, Bhagalpur, Bihar, India

²Tutor, Department of Microbiology, JLNMCH, Bhagalpur, Bihar, India

³Assistant Professor, Department of Pediatrics, JLNMCH, Bhagalpur, Bihar, India

Received: 24-11-2021 / Revised: 22-12-2021 / Accepted: 07-01-2022

Abstract

Introduction: Metallo-beta lactamase production is the most common mechanism of carbapenem resistance. Metallo-beta lactamase is a zinc dependent enzyme belonging to Ambler class B that can hydrolyse all beta lactam antibiotics including carbapenem. Keeping it in mind we have conducted a research to find out MBL positivity rate in clinical isolates of Pseudomonas in a tertiary care hospital of Bihar. **Methodology:** This is a descriptive cross-sectional study conducted in the Department of Microbiology, Jawaharlal Nehru Medical College and Hospital, Bhagalpur, Bihar, India over a period of 1 year from May 2020 to April 2021. Depending upon site of infection samples were collected like urine, pus, sputum, BAL, ear swab etc in sterile container. Carbapenem resistance was suspected when either imipenem or meropenem was resistant (MIC > 8 µg/ml). MBL production was tested in all carbapenem resistant Pseudomonas species by Combined disc test with imipenem and was confirmed by MBL- E test. **Results:** A total of 1357 various body fluids were obtained from various wards of the hospital over the study period, out of which, Pseudomonas species were isolated from 207 samples. Among 207 samples, 181 were P. aeruginosa and 26 were P. putida. Majority of the samples were obtained from OPD followed by IPD and ICU. MBL production (MBL +) was noted among 13.5% (28/207) of isolates and carbapenem resistance was observed among 14.9% (31/207). When both these characters were matched, it was noted that almost two-third (74.2%) of carbapenem resistance (CR) Pseudomonas were MBL positive. Both these characteristics were more common among P. putida as compared with P. aeruginosa. **Conclusion:** MBL producing Pseudomonas is difficult to treat but easy to prevent by proper hospital infection control measures and antibiotic policy.

Key Words: Metallo-Beta Lactamase, Pseudomonas aeruginosa Species

This is an Open Access article that uses a funding model which does not charge readers or their institutions for access and distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>) and the Budapest Open Access Initiative (<http://www.budapestopenaccessinitiative.org/read>), which permit unrestricted use, distribution, and reproduction in any medium, provided original work is properly credited.

Introduction

Pseudomonas is the gram-negative bacilli that are strict aerobes, motile with one or two flagella, utilise glucose oxidatively, and are oxidase positive. It belongs to Pseudomonadaceae family and in molecular taxonomy to fluorescent group of r RNA group I[1]. Another member of this Fluorescent group are Pseudomonas fluorescence and Pseudomonas putida that are rarely involved in clinical diseases in human[2]. With the help of different virulent factors like pyocyanin, exotoxin A, exoenzyme S, protease, phospholipase, rhamnolipids they produce both community acquired infections like otitis externa, keratitis, varicose vein ulcer and hospital acquired infection like Catheter associated urinary tract infection (CAUTI), Ventilator associated pneumonia (VAP), burn infection, bedsores, septicemia and necrotising pneumonia in cystic fibrosis patients etc[2]. They have highly evolved Quorum sensing mechanism by which they can easily form biofilm and prevent attack of antibiotic[2].

Most important factor that makes it so much dominant in hospital environment that it can resist or even can utilise some disinfectants/antiseptics like cetrimide for their nutrition[1], so they easily grow in hospital environment and ICU. They are responsible for 10% of all hospital acquired infection[2]. Aminoglycoside (gentamicin, amikacin, tobramycin), antipseudomonal penicillin (piperacillin, ticarcillin) and cephalosporin like Cefazidime are used to treat Pseudomonas infection but resistance against these antibiotics are common today[2].

*Correspondence

Dr. Sagar Kumar

Tutor, Department of Microbiology, JLNMCH, Bhagalpur, Bihar, India

E-mail: dr.sagarkr@gmail.com

Beta lactamase destroy beta lactam ring of antibiotic and make them ineffective against Pseudomonas[3]. carbapenem is the drug of choice in extended spectrum beta lactamase producing Pseudomonas[3]. This was derived from thienamycin, a naturally derived product of Streptomyces cattleya[4]. Ertopenem, Doripenem, imipenem, Meropenem and Faropenem are example of carbapenem but imipenem and Meropenem are most commonly used carbapenem in India[4]. With the progress of time irrational and inappropriate use of carbapenem led to emergence of carbapenem resistant Pseudomonas first in Japan in 1991[5] and then in different part of the world. In India first case of MBL producing Pseudomonas was reported in 2002[6, 7]. Mechanism of carbapenem resistance is mainly three types, first due to increase expression of porin in cell wall, second due to increase activity of efflux pump and third- production of metallo-beta lactamase[7]. Metallo-beta lactamase production is the most common mechanism of carbapenem resistance[7]. Metallo-beta lactamase is a zinc dependent enzyme belonging to Ambler class B that can hydrolyse all beta lactam antibiotics including carbapenem[8]. Ambler class A, C, D beta lactamases use serine as active site so they can be easily degraded by beta lactamase inhibitor like clavulanic acid or sulbactam. But metallo-beta lactamase cannot be inhibited by clavulanic acid or sulbactam so MBL producing Pseudomonas is now emerging as a nightmare for treating physician. Besides that, resistance determinant of MBL is located in highly mobile genetic element allowing easy dissemination from patient to patient or even from patient to health care providers[2]. So, prevention is the always better option than treatment of MBL Pseudomonas infection. Clinician in every hospital should know the local prevalence of MBL producing Pseudomonas to formulate proper antibiotic policy and hospital infection control strategy to prevent outbreak of this dangerous superbug.

Keeping it in mind we have conducted a research to find out MBL positivity rate in clinical isolates of Pseudomonas in a tertiary care hospital of Bihar.

Methodology

This is a descriptive cross-sectional study conducted in the Department of Microbiology, Jawaharlal Nehru Medical College and Hospital, Bhagalpur, Bihar, India over a period of 1 year from May 2020 to April 2021. Depending upon site of infection samples were collected like urine, pus, sputum, BAL, ear swab etc in sterile container.

Sample Processing

All samples were inoculated immediately into Blood agar and MacConkey agar media (HiMedia, Mumbai) and incubated for 18-24 hrs. At 37°C in incubator. Next day growth was observed, and gram stain was performed. All the positive growth which was oxidase positive selected and put in Vitek[2] identification system (Biomerieux). Identification and antibiogram of oxidase positive growth was done in fully automated Vitek[2]. Carbapenem resistance was suspected when either imipenem or meropenem was resistant (mic>8 µl/ml). MBL production was tested in all carbapenem resistant Pseudomonas species by Combined disc test with imipenem and was confirmed by MBL- E test.

Combined Disc Test[5]

Two 10 µg IPM disks were put on the MHA plate seeded with the test organism. 10 µL of EDTA solution (750 µg) was added to one of them. The plate was incubated for 16-18 hrs at 35°C. If the increase in inhibition zone with the IPM + EDTA disk was >7 mm than the IPM disk alone, it was suspected as MBL positive.

MBL Epsilon meter Test (E-Test)[5]

The MBL E-strip with seven-dilution range of IPM (4-256 µg/mL) in one side and IPM plus EDTA (1-64 µg/mL) on another side was put on MHA plate seeded with test organism. The plate was then incubated in incubator at 35°C for 18-20 hrs. MIC ratio of IPM/IPM + EDTA of >8, or reduction of IPM MIC by >3log2 dilutions in the presence of EDTA confirmed MBL production.

Results

A total of 1357 various body fluids were obtained from various wards of the hospital over the study period, out of which, Pseudomonas species were isolated from 207 samples. Among which 172 were from urine, 21 were from pus and rest 14 were from another sites like sputum, ear swab, bronchoalveolar lavage (BAL), endotracheal tube (ET) aspirate etc. Among 207 samples, 181 were P. aeruginosa and 26 were P. putida. Majority of the samples were obtained from OPD followed by IPD and ICU.

MBL production (MBL +) was noted among 13.5% (28/207) of isolates and carbapenem resistance was observed among 14.9% (31/207). When both these characters were matched, it was noted that almost two-third (74.2%) of carbapenem resistance (CR) Pseudomonas were MBL positive. Both these characteristics were more common among P. putida as compared with P. aeruginosa. [Figure 1] Both MBL producing strains were more common among males than females (19:9). Majority of the MBL positive organism were isolated from the patients with age group of 40-60 years. Various factors affecting CR and MBL + has been shown in table 1 and 2. Antibiogram of MBL positive and MBL negative Pseudomonas species was compared in table 3.

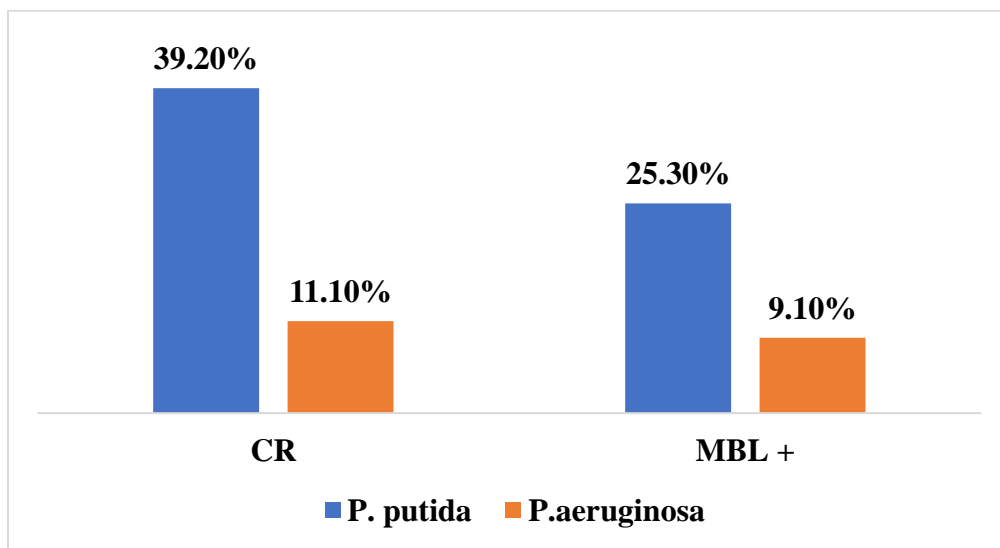


Figure 1: Carbapenem resistance and MBL Positivity Rate in both the species of Pseudomonas

Table 1: Sample wise and ward wise distribution of carbapenem resistance

Factor	Classification	Carbapenem resistance	MBL +
Type of sample	Urine	9.3%	9.1%
	Pus	34.2%	27.3%
	Others	8.9%	1.1%
Point of collection	OPD	11.1%	7.5%
	IPD	13.4%	11.2%
	ICU	24.5%	17.3%

Table 2: Antibiogram showing sensitivity of both the species to various antibiotics

Antibiotic	Sensitivity of MBL + isolates (%)	Sensitivity of MBL – isolates (%)
Ampicillin	7.2	42.1
Amoxicillin-clavulanic acid	20.2	67.5
Piperacillin-Tazobactam	40.2	91.4
Cefuroxime	2.1	44.7

Ceftazidime	20.2	67.5
Cefoperazone-sulbactam	40.2	91.4
Cefepime	2.1	66.9
Amikacin	22.1	100
Gentamycin	20.2	82.1
Ciprofloxacin	27.8	67.5
Tigecycline	12.5	82.1
Nitrofurantoin	15.4	75
Colistin	100	100
Trimethoprim/Sulfamethoxazole	20.2	44.7

Discussion

Pseudomonas has the ability to grow and multiply in moist environment and equipment including sinks, drain flower vas, hydrotherapy pools, ponds, river and even in distilled water[2]. In hospital setting it can grow in many disinfectants and pharmaceutical product posing serious problem in infection control[2]. Blue green colour pyocyanin produced by *P. aeruginosa* and pyoverdine produced by another *Pseudomonas* act as virulent factor. With the help of metallo-beta lactamase enzyme they become antibiotic resistant[9]. Lots of phenotypic tests are available for detection of metallo-beta lactamase production in *Pseudomonas*. Principal of all those test is the ability of metal ion chelator like EDTA or thiol compound to inhibit activity of MBL[10]. Among all these tests CDT showed good sensitivity (79%) than other tests like DDST (70.8%) and Disc potentiation test (54.2%) in a study done by Ranjan et al[5]. Another studies like Samuelson et al[11], Qu et al[12], Biradar et al[13] also had reported that CDT was better than all other tests. It was easy to perform and cheaper and having objective interpretation[5]. Rit et al[14] in Kolkata had reported that CDT was having same accuracy with MIC detection. So we had selected CDT as screening test for MBL detection in our study. CDT could be performed using ceftazidime or imipenem but as *Pseudomonas* might have other resistance mechanism other than MBL production to ceftazidime[15] we had used imipenem for CDT. For confirmation, PCR analysis of MBL gene was the gold standard but it was not feasible in routine microbiology laboratory in a developing country like India[5]. In contrast, MBL E test showed good specificity (98%) in study by Khosravi et al[16], Walsh et al[17] and Segal et al[18]. So, we had selected MBL E test as the confirmatory test for MBL detection in our study.

In our study among 207 isolated *Pseudomonas* strains, 14.9% were carbapenem resistant which was slight higher than Kanungo et al[19] (10.9%) but lower than many others[3, 5, 9, 10, 13, 20]. Implementation of strict antibiotic policy in our hospital might be responsible for such low level of carbapenem resistance in our study. Among carbapenem resistant *Pseudomonas*, 74.2% were MBL producing in our study which was almost same as that of study done by Biradar et al[13] (74%) but was lower than some others[10, 21-24]. In our study MBL positivity rate was 13.5%. This is lower than many previous researchers[3, 8-10, 12-16, 20, 30-36]. Variation in MBL positivity rate in different studies might be due to several factors like Geographical area, Infection control attitude of the hospital, sample size and method of testing. But overall in our hospital MBL positivity rate was lower than most of the studies in India and it should be maintained in future by strict hospital infection control and antibiotic policy. Slight male preponderance was seen in MBL positive *Pseudomonas* infection, but it was not significant. There are studies reporting male preponderance[5, 9, 13] and at the same time some have reported female dominance[37]. Highest no of MBL positive isolates came from older age group (>40) in our study. Choudhary et al[9] and Biradar et al[13] had reported middle age group (31-60) was the most commonly affected age group. Long hospital stay, frequent hospital admission due to age related problems and relative immunocompromised status of older age group[5] might be responsible for higher MBL positivity rate in older age group in our study.

MBL positivity in *Pseudomonas* was highest in pus isolates followed by urine that is similar to others[3, 9, 10, 13, 21]. Wound easily

comes in contact of hospital environment leading to easy colonisation compared to bladder that requires catheter manipulation to get infected by *Pseudomonas*. MBL positivity of *Pseudomonas* in our study was highest in ICU compared to IPD and OPD. It was consistent with the study by Kaur et al[8] but not with the study done by Easwaran et al[4] where MBL positivity was highest in IPD followed by ICU. More number of invasive interventions prolong stay in ICU, serious nature of the disease in ICU patients all might be responsible for high MBL positivity rate in ICU. Most of the study done in India about MBL production was in *Pseudomonas aeruginosa* which was the most common species of *Pseudomonas*. As per our literature search no data was available about MBL positivity in *Pseudomonas putida* in India. MBL positivity was higher in *P. putida* than *P. aeruginosa* in our study. It was in consistent with a Spanish study[38] where 14% of *P. putida* and 0.3% of *P. aeruginosa* was MBL positive. *P. putida* acts as environmental reservoir of MBL resistance gene and acts as a donor of this gene to *P. aeruginosa*.

In antibiotic sensitivity test, MBL positive *Pseudomonas* species showed poor sensitivity against most of the antibiotics like ampicillin, amoxiclav, Cefuroxime, Ceftazidime, Cefoperazone sulbactam, amikacin, gentamicin, ciprofloxacin, cotrimoxazole, Nitrofurantoin compared to MBL negative strains. This finding was consistent with another studies[9, 10, 13]. In every hospital MBL positivity in *Pseudomonas* should be checked by Microbiologist as routine laboratory practice and local prevalence of that superbug should be kept in mind during hospital infection control policy making to prevent outbreak of this highly communicable resistance determinant.

Conclusion

MBL producing *Pseudomonas* is difficult to treat but easy to prevent by proper hospital infection control measures and antibiotic policy. In our study MBL positivity rate (12.5%) in *Pseudomonas* was lower when compared to most of the similar studies in India. MBL prevalence in *Pseudomonas putida* was higher than *Pseudomonas aeruginosa*. Colistin was the only antibiotic with good sensitivity against this dangerous superbug.

Conflict of interest

None declared by any of the authors

References

- Koneman EW, Allen SD, Janda WM, et al. Color Atlas and Textbook of Diagnostic Microbiology. 6th edn. Philadelphia, USA: Lippincott Raven Publishers 1997.
- Greenwood D. Medical microbiology. 18th edn. Edinburgh: Churchill Livingstone/Elsevier 2012.
- Rajput A, Prajapati B, Chauhan B, et al. Prevalence of Metallo-Betalactamases (MBL) producing *Pseudomonas aeruginosa* in a tertiary care hospital. Ind J Basic & App Med Res 2012; 1(4):304-8.
- Easwaran S, Ramasamy R. Prevalence of metallo β lactamases producing *Pseudomonas* spp. and *Acinetobacter* spp. in a tertiary care teaching hospital. J Drug Discovery Ther 2017; 5(7):35-9.
- Ranjan S, Banashankari GS, Babu PR. Evaluation of phenotypic tests and screening markers for detection of metallo- β -lactamases in clinical isolates of *Pseudomonas aeruginosa*: a prospective study. Medical Journal of Dr. DY Patil University 2015; 8(5):599-605.
- Peleg AY, Franklin C, Bell JM, et al. Dissemination of the metallo- β -lactamase gene bla_{IMP-4} among gramnegative

- pathogens in a clinical setting in Australia. *Clinical Infectious Diseases* 2005; 41(11):1549-56.
7. Nandi A, Bhattacharya S, Biswas S, et al. A study on Metallo- β lactamase producing imipenem nonsusceptible multi-drug resistant *Pseudomonas aeruginosa* in different clinical specimens in a tertiary care hospital in Kolkata. *J Dent Med Sci* 2014; 13(6):13-7.
 8. Kaur A, Singh S. Prevalence of Extended Spectrum Betalactamase (ESBL) and Metallobetalactamase (MBL) producing *Pseudomonas aeruginosa* and *acinetobacter baumannii* isolated from various clinical samples. *Journal of Pathogens* 2018; 2018:6845985.
 9. Choudhary V, Pal N, Hooja S. Prevalence and antibiotic resistance pattern of Metallo- β -lactamase-producing *Pseudomonas aeruginosa* isolates from clinical specimens in a tertiary care hospital. *Journal of Mahatma Gandhi Institute of Medical Sciences* 2019; 24(1):19-22.
 10. Mishra SN, Biswal SR, Behera BK, et al. Detection of prevalence of metallo-beta lactamases in clinical isolates of imipenem resistant *Pseudomonas aeruginosa* from neonatal septicemia cases in a tertiary hospital in Odisha, India. *Int J Contemp Pediatr* 2018; 5(1):61-6.
 11. Samuelsen O, Buaro L, Giske CG, et al. Evaluation of phenotypic tests for the detection of metallo- β -lactamase-producing *Pseudomonas aeruginosa* in a low prevalence country. *Journal of Antimicrobial Chemotherapy* 2008; 61(4):827-30.
 12. Qu TT, Zhang JL, Wang J, et al. Evaluation of phenotypic tests for detection of Metallo- β -lactamase-producing *Pseudomonas aeruginosa* strains in China. *Journal of Clinical Microbiology* 2009; 47(4):1136-42.
 13. Biradar S, Roopa C. Prevalence of Metallo-beta lactamase producing *Pseudomonas aeruginosa* and its antibiogram in a tertiary care centre. *Int J Curr Microbiol Appl Sci* 2015; 4(9):952-6.
 14. Rit K, Chakraborty B, Dey R, et al. Prevalence of *Pseudomonas aeruginosa* and *acinetobacter* spp. producing metallo- β -lactamase in a tertiary care hospital. *Journal of Dr. NTR University of Health Sciences* 2013; 2(1):18-21.
 15. Behera B, Mathur P, Das A, et al. An evaluation of four different phenotypic techniques for detection of metallo- β -lactamase producing *Pseudomonas aeruginosa*. *Indian Journal of Medical Microbiology* 2008; 26(3):233-7.
 16. Khosravi Y, Loke MF, Chua EG, et al. Phenotypic detection of metallo- β -lactamase in imipenem-resistant *Pseudomonas aeruginosa*. *The Scientific World Journal* 2012; 2012:654939.
 17. Walsh TR, Bolmström A, Qwärnström A, et al. Evaluation of a new Etest for detecting metallo- β -lactamases in routine clinical testing. *Journal of Clinical Microbiology* 2002; 40(8):2755-9.
 18. Segal H, Elisha BG. Use of Etest MBL strips for the detection of carbapenemases in *Acinetobacter baumannii*. *Journal of Antimicrobial Chemotherapy* 2005; 56(3):598.
 19. Shashikala, Kanungo R, Srinivasan S, et al. Emerging resistance to carbapenems in hospital acquired *Pseudomonas* infection: a cause for concern. *Indian Journal of Pharmacology* 2006; 38(4):287-8.
 20. Varaiya A, Kulkarni M, Bhalekar P, et al. Incidence of metallo-beta-lactamase-producing *Pseudomonas aeruginosa* in diabetes and cancer patients. *Indian Journal of Pathology and Microbiology* 2008; 51(2):200-3.
 21. Chand AE, Chauhan PS, Sharma S, et al. Prevalence of Metallo-beta-lactamase production in imipenem-resistant *Pseudomonas* in tertiary care center at Kota region. *Int J Sci Stud* 2016; 4(3):87-91.
 22. Attal RO, Basak S, Mallick SK, et al. Metallo-betalactamase producing *Pseudomonas aeruginosa*: an emerging threat to clinicians. *J Clin Diagn Res* 2010; 4:2691-6.
 23. Fam N, Diab M, Helmi H, et al. Phenotypic detection of metallo- β -Lactamases and extended spectrum β -Lactamases among Gram-negative bacterial clinical isolates. *Egyptian Journal of Medical Microbiology* 2006; 15(4):719-30.
 24. Irfan S, Zafar A, Guhar D, et al. Metallo- β -lactamase-producing clinical isolates of *acinetobacter* species and *Pseudomonas aeruginosa* from intensive care unit patients of a tertiary care hospital. *Indian Journal of Medical Microbiology* 2008; 26(3):243-5.
 25. Navaneeth BV, Sridaran D, Sahay D, et al. A preliminary study on metallo-[beta]-lactamase producing *Pseudomonas aeruginosa* in hospitalized patients. *Indian Journal of Medical Research* 2002; 116:264-7.
 26. Hemalatha V, Sekar U, Kamat V. Detection of metallo- β -lactamase producing *Pseudomonas aeruginosa* in hospitalized patients. *Indian J Med Res* 2005; 122(2):148-52.
 27. Castanheira M, Bell JM, Turnidge JD, et al. Carbapenem resistance among *Pseudomonas aeruginosa* strains from India: evidence for nationwide endemicity of multiple metallo- β -lactamase clones (VIM-2,-5,-6, and-11 and the newly characterized VIM-18). *Antimicrobial Agents And Chemotherapy* 2009; 53(3):1225-7.
 28. Owlia P, Saderi H, Karimi Z, et al. Phenotypic detection of Metallo-beta-Lactamase producing *Pseudomonas aeruginosa* strains isolated from burned patients. *Iranian Journal of Pathology* 2008; 3(1):20-5.
 29. Manoharan A, Chatterjee S, Mathai D, et al. Detection and characterization of metallo-beta lactamases producing *Pseudomonas aeruginosa*. *Indian Journal of Medical Microbiology* 2010; 28(3):241-4.
 30. Kumar SH, De Anuradha S, Baveja SM, et al. Prevalence and risk factors of metallo β -lactamase producing *Pseudomonas aeruginosa* and *acinetobacter* species in burns and surgical wards in a tertiary care hospital. *Journal of Laboratory Physicians* 2012; 4(1):39-42.
 31. Kali A, Srirangaraj SK, Kumar S, et al. Detection of metallo-beta-lactamase producing *Pseudomonas aeruginosa* in intensive care units. *The Australasian Medical Journal* 2013; 6(12):686-93.
 32. Mendiratta DK, Deotale V, Narang P. Metallo-betalactamase producing *Pseudomonas aeruginosa* in a hospital from a rural area. *Indian Journal of Medical Research* 2005; 121(5):701-3.
 33. Agrawal G, Lodhi RB, Kamalakar UP, et al. Study of metallo-beta-lactamases production in clinical isolates of *Pseudomonas aeruginosa*. *Indian J Med Microbiol* 2008; 26(4):349-51.
 34. Pitout JDD, Gregson DB, Poirel L, et al. Detection of *Pseudomonas aeruginosa* producing metallo- β -lactamases in a large centralized laboratory. *Journal of Clinical Microbiology* 2005; 43(7):3129-35.
 35. Ibukun A, Tochukwu N, Tolu O. Occurrence of ESBL and MBL in clinical isolates of *Pseudomonas aeruginosa* from Lagos, Nigeria. *J Am Sci* 2007; 3(4):81-5.
 36. Chaudhary U, Bhaskar H, Sharma M. Imipenem-EDTA disk method for rapid identification of metallo-betalactamase producing Gram-negative bacteria. *Indian Journal of Medical Research* 2008; 127(4):406-7.
 37. Anil C, Shahid RM. Antimicrobial susceptibility patterns of *Pseudomonas aeruginosa* clinical isolates at a tertiary care hospital in Kathmandu, Nepal. *Asian J Pharm Clin Res* 2020; 6(3):235-8.
 38. Juan C, Zamorano L, Mena A, et al. Metallo- β -lactamase-producing *Pseudomonas putida* as a reservoir of multidrug resistance elements that can be transferred to successful *Pseudomonas aeruginosa* clones. *Journal of Antimicrobial Chemotherapy* 2021; 65(3):474-8.